

## RAPD Protocol

Protocol for Promega and Stoffel brand Taq enzymes (1 reaction):	Protocol for Gibco brand Taq enzyme (1 reaction):	Final Conc.
<p>4 <math>\mu</math>l dNTP (5mM) mix 2.5 <math>\mu</math>l 10X PCR buffer 8.33 <math>\mu</math>l H<sub>2</sub>O</p> <hr/> <p>5 <math>\mu</math>l (25 mM) MgCl<sub>2</sub> 0.22 <math>\mu</math>l (5 U/<math>\mu</math>l) Taq</p> <hr/> <p>3 <math>\mu</math>l primer (10 ng/<math>\mu</math>l) 3 <math>\mu</math>l template DNA (10 ng/<math>\mu</math>l)</p> <hr/> <p>Total vol = 26.05 <math>\mu</math>L 3 <math>\mu</math>l loading dye</p>	<p>4 <math>\mu</math>l dNTP (5mM) mix 2.5 <math>\mu</math>l 10X PCR buffer 10.83 <math>\mu</math>l H<sub>2</sub>O</p> <hr/> <p>2.5 <math>\mu</math>l (50 mM) MgCl<sub>2</sub> 0.22 <math>\mu</math>l (5 U/<math>\mu</math>l) Taq</p> <hr/> <p>3 <math>\mu</math>l primer (10 ng/<math>\mu</math>l) 3 <math>\mu</math>l template DNA (10 ng/<math>\mu</math>l)</p> <hr/> <p>Total vol = 26.05 <math>\mu</math>L 3 <math>\mu</math>l loading dye</p>	<p>0.8 mM dNTP mix (0.2 mM each dNTP)</p> <p>5 mM MgCl<sub>2</sub> 0.042 U/<math>\mu</math>l Taq</p> <p>1.15 ng/<math>\mu</math>l primer 1.15 ng/<math>\mu</math>l DNA</p>
<p><b>dNTP mix (5mM):</b> 12.5 <math>\mu</math>l of 100mM dATP 12.5 <math>\mu</math>l of 100mM dGTP 12.5 <math>\mu</math>l of 100mM dCTP 12.5 <math>\mu</math>l of 100mM dTTP</p> <hr/> <p>Add water up to 1000 <math>\mu</math>l</p> <p><b>Loading buffer (6X):</b> 30% Glycerol (make 25ml with H<sub>2</sub>) 0.25% Bromophenol blue 0.25% Xylene cyanole</p> <hr/> <p>In a cylinder, cover with parafilm and mix by inverting.</p>	<p><b>Ladder mix:</b> 100 bp ladder 50 <math>\mu</math>g; 1 <math>\mu</math>g/<math>\mu</math>l 166 <math>\mu</math>l Loading Buffer 784 <math>\mu</math>l H<sub>2</sub>O</p> <hr/> <p>total of 100 <math>\mu</math>l</p> <p><b>PCR profile:</b> 94°C 1 min ———— 35°C 1 min ———— 3 cycles 72°C 2 min ———— 94°C 10s ———— 40°C 20s ———— 34 cycles 72°C 2 min ———— 72°C 5 min 4°C soak</p>	